

## 柴达木盆地唐古特白刺种子的化学成分研究

王洪伦<sup>1</sup>, 李玉林<sup>1,2</sup>, 王小艳<sup>1,2</sup>, 索有瑞<sup>1\*</sup>

<sup>1</sup>中国科学院西北高原生物研究所, 西宁 810001; <sup>2</sup>中国科学院研究生院, 北京 100039

**摘要:**从唐古特白刺种子 75%乙醇提取物中分离得到 6 种化合物,应用波谱方法及与文献对照的手段鉴定为:胡萝卜素 (daucosterol, 1), 4-羟基吡咯羧酸 (2), 槲皮素 (quercetin, 3), 尿囊素 (allantoin, 4), 1, 2, 3, 4-四氢-1-甲基-β-羧oline-3-羧基酸 (5), L-tyrosine (6)。除槲皮素外的其他五种化合物均为首次从该植物中分离得到。

**关键词:**唐古特白刺; 种子; 化学成分; 尿囊素; 分离; 结构鉴定

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### Chemical Constituents of *Nitraria tangutorum* Seed from Qaidam Basin

WANG Hong-lun<sup>1</sup>, LI Yu-lin<sup>1,2</sup>, WANG Xiao-yan<sup>1,2</sup>, SUO You-rui<sup>1\*</sup>

<sup>1</sup>Northwest Institute of Plateau Biology, Chinese Academy of Sciences, Xining 810001, China;

<sup>2</sup>Graduate University of the Chinese Academy of Sciences, Beijing 100039, China

**Abstract:** Six compounds were isolated from the 75% ethanol extract of *Nitraria tangutorum* seed. On the basis of spectroscopic methods including <sup>1</sup>H NMR, <sup>13</sup>C NMR and ES/MS and comparison with literature, their structures were elucidated as daucosterol (1), 4-hydroxyisochlorogenic acid (2), quercetin (3), allantoin (4), 1, 2, 3, 4-tetrahydro-1-methyl-β-carboline-3-carboxylic acid (5) and L-tyrosine (6). Compounds 1, 2, 3, 5 and 6 were isolated from *Nitraria tangutorum* for the first time.

**Key words:** *Nitraria tangutorum*; seed; 1, 2, 3, 4-tetrahydro-1-methyl-β-carboline-3-carboxylic acid; allantoin

### Introduction

The genus *Nitraria* (Zygophyllaceae) is a shrub that bears edible berries and widely distributed in the Middle East, Central Asia, and the Northwest region of China. Among the *Nitraria* species, only *N. tangutorum* Bobr grows in China, especially in the desert of Qinghai-Tibetan Plateau. A main function of a *N. tangutorum* Bobr forest is to conserve the soil and water from the wind-blown sand<sup>[1,2]</sup>. In addition, its leaves, fruits and seeds are often used in folk medicines such as antispasmodic, antineuropathic, and anti-arrhythmic agent<sup>[3,4]</sup> to cure weaknesses in the spleen and stomach<sup>[5,6]</sup> and decrease blood lipid levels and anti-oxidation<sup>[7]</sup>. The chemical constituents of *Nitraria tangutorum* seed collected from Gansu Province and the chemical con-

stituents of *Nitraria tangutorum* leaves from Ningxia Province had been reported<sup>[5,8]</sup>. And it revealed the presence of flavonoids, phenolic acids and alkaloids. However, the chemical constituents of *Nitraria tangutorum* seeds from Qaidam basin, Qinghai Province were not reported. In this study, six compounds were isolated from the ethanol extract of the *Nitraria tangutorum* seed collected from Qaidam basin. Their structures were elucidated as daucosterol (1), 4-hydroxyisochlorogenic acid (2), quercetin (3), allantoin (4), 1, 2, 3, 4-tetrahydro-1-methyl-β-carboline-3-carboxylic acid (5), L-tyrosine (6) on the basis of spectral data or in comparison with literature.

### Materials and Methods

#### Apparatus and materials

All melting points were determined on a PHMK micro-melting point apparatus and uncorrected IR spectra were recorded on a Nicolet NEXUS 670 FT-IR spectrophotometer (KBr). NMR spectra were obtained on a

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\* Corresponding author. Tel: 86-971-6143857; E-mail: yrsuo@nwipb.ac.cn

Bruker AM400 and Varian Mercury 400B NMR spectrometer Elements were analyzed on the Elementar Analysensysteme GmbH VarioEL. ESHMS were carried on a Bruker Apex . Silica gel (Qingdao Haiyang Chemical Co., Ltd., China), RP-C18 and Sephadex LH20 (Pharmacia Co., American) were used for column chromatography. The *Nitria tangutonum* seeds were collected in September 2003 in Dulan county, Qinghai Province and identified by Prof Liu Shang-wu.

### Extraction and isolation

The air-dried and powdered seeds (15 kg) of *Nitria tangutonum* were extracted firstly by supercritical carbon dioxide for extracting the fatty acids, and then the seeds were extracted by 75% EtOH three times under reflux. After the solvent was evaporated *in vacuo*, the residue (1200 g) was suspended in water and extracted with petroleum ether (bp. 60-90 °C), chloroform, ethyl acetate and *n*-butanol respectively. The ethyl acetate fraction (45 g) was separated over silica column chromatography (CC) eluting with chloroform-methanol (70:1:0:1) to obtain compounds **1** (30 mg) and **3** (39 mg). The *n*-butanol (75 g) fraction was isolated on silica CC eluting with chloroform-methanol (50:1:0:1), and 5 fractions (Fr1-Fr5) were obtained according to detection by TLC (GF<sub>254</sub>). Fr1 was separated by silica gel CC and purified Sephadex LH-20 column eluting methanol to obtain compound **4** (115 mg). Fr3 was purified by Sephadex LH-20 and RP-C18 to give compound **5** (43 mg) and compound **6** (16 mg). Fr5 was isolated on silica gel CC eluting with chloroform-methanol (4:1:0:1) to yield compound **2** (220 mg).

### Identification

**Daucosterol (1)** White powder (methanol), mp. 270-272 °C, C<sub>35</sub>H<sub>60</sub>O<sub>6</sub>. <sup>1</sup>H NMR (Pyridine-*d*<sub>5</sub>, 400 MHz): 5.34 (1H, m, H-6), 5.05 (1H, d, *J* = 7.6 Hz, H-1), 4.60-3.90 (m, H-2-6), 2.80-1.00 (m), 0.97-0.83 (m, 5 ×-CH<sub>3</sub>), 0.64 (s, -CH<sub>3</sub>); <sup>13</sup>C NMR (Pyridine-*d*<sub>5</sub>, 100 MHz): 149.9, 121.9, 102.6, 78.6, 78.3, 78.1, 75.3, 71.7, 62.8, 56.8, 56.2, 50.3, 46.0, 42.5, 39.9, 39.3, 37.5, 36.9, 36.4, 34.2, 32.2, 32.0, 30.2, 29.4, 28.5, 26.3, 24.5, 23.4, 21.3, 20.0, 19.4, 19.2, 19.0, 12.1, 12.0. Its data were con-

sistent with those of daucosterol<sup>[9]</sup>.

**4-Hydroxypipelic acid (2)** White powder (EtOH), mp. 261-263 °C, C<sub>6</sub>H<sub>11</sub>NO<sub>3</sub>. Elements analysis (%): C 46.42, H 7.05, N 7.75;  $R_{\max}^{KB}$  cm<sup>-1</sup>: 3266, 3121, 2938, 1610. <sup>1</sup>H NMR (D<sub>2</sub>O, 400 MHz): 4.05 (1H, m, H-4), 3.73 (1H, dd, *J* = 11.6, 3.6 Hz, H-2), 3.12 (2H, m, H-6), 2.04, 1.79 (1H, m, H-4), 1.72 (2H, m, H-5); <sup>13</sup>C NMR (D<sub>2</sub>O, 100 MHz): 177.04 (COOH), 64.64 (C-4), 56.74 (C-2), 41.35 (C-6), 35.61 (C-3), 30.74 (C-5). The  $R_{\max}$  NMR data were identical to those reported<sup>[10]</sup>.

**Quercetin (3)** Yellow powder (methanol), mp. 310-312 °C, C<sub>15</sub>H<sub>10</sub>O<sub>7</sub>. <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz): 7.72 (1H, d, *J* = 2.0 Hz, H-2), 7.62 (1H, dd, *J* = 8.4, 2.0 Hz, H-6), 6.87 (1H, d, *J* = 8.4 Hz, H-5), 6.38 (1H, d, *J* = 2.0 Hz, H-8), 6.17 (1H, d, *J* = 2.0 Hz, H-6); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 100 MHz): 177.3 (C-4), 165.6 (C-7), 162.5 (C-9), 158.2 (C-5), 148.8 (C-4), 148.0 (C-2), 146.2 (C-3), 136.6 (C-3), 124.1 (C-1), 121.6 (C-6), 116.2 (C-5), 116.0 (C-2s), 104.5 (C-10), 99.2 (C-6), 94.4 (C-8). Its NMR data were consistent with those of quercetin<sup>[11]</sup>.

**Allantoin (4)** White powder (methanol), mp. 237-239 °C, C<sub>4</sub>H<sub>6</sub>N<sub>4</sub>O<sub>3</sub>, HR-ESHMS: [M + H]<sup>+</sup> 159.0513.  $R_{\max}^{KB}$  cm<sup>-1</sup>: 3438, 3345, 1782, 1709, 1658, 1603, 1530, 1184. <sup>1</sup>H NMR (DMSO, 400 MHz): 10.53 (1H, s), 8.05 (1H, s), 6.88 (1H, d, *J* = 8.0 Hz), 5.78 (2H, s), 5.24 (1H, d, *J* = 6.8 Hz), 5.23 (1H, s); <sup>13</sup>C NMR (DMSO, 100 MHz): 173.6 (-NH-CO-NH<sub>2</sub>), 157.4 (-NH-CO-), 156.8 (-NH-CO-), 62.4 (CH). Its melting point and *R<sub>f</sub>* were same to an authentic allantoin sample (National Institute for the Control of Pharmaceutical and Biological Products, No. 1501-200001). It was identified as allantoin<sup>[12,13]</sup>.

**1, 2, 3, 4-Tetrahydro-1-methyl- $\beta$ -carboline-3-carboxylic acid (5)** White powder (methanol), mp. 286-288 °C. C<sub>13</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>, HR-ESHMS: [M + H]<sup>+</sup> 231.1140.  $R_{\max}^{KB}$  cm<sup>-1</sup>: 3294, 2981, 2922, 1644, 1577, 1384, 742; <sup>1</sup>H NMR (DMSO, 400 MHz): 11.12 (1H, s), 7.43 (1H, d, *J* = 8.0 Hz, H-9), 7.33 (1H, d, *J* = 8.0 Hz, H-12), 7.08 (1H, t, *J* = 7.2, 7.6 Hz, H-11), 7.00 (1H, t, *J* = 7.6, 7.2 Hz, H-10), 4.56 (1H, m, H-3), 3.69 (1H, dd, *J* = 12.0, 4.8 Hz, H-5), 3.22, 2.80

(2H, m, H-6), 1.61 (3H, d,  $J = 6.8$  Hz, -CH<sub>3</sub>); <sup>13</sup>C NMR (DMSO, 100 MHz): 170.45 (-COOH), 136.87 (C-13), 131.73 (C-2), 126.33 (C-8), 122.08 (C-11), 119.52 (C-10), 118.52 (C-9), 111.77 (C-12), 106.93 (C-7), 58.05 (C-5), 49.63 (C-3), 23.23 (C-6), 17.03 (-CH<sub>3</sub>). The IR, MS, NMR spectral data were identical with those of 1, 2, 3, 4-tetrahydro-1-methyl- $\beta$ -carboline-3-carboxylic acid.

**L-Tyrosine (6)** White powder (methanol), mp. 244-246. IR  $\text{KBr cm}^{-1}$ : 3206, 2928, 1611, 1588, 1513, 1454, 1329, 1245; <sup>1</sup>H NMR (DMSO + D<sub>2</sub>O, 400 MHz): 7.02 (2H, d,  $J = 8.0$  Hz, H-6, 8), 6.66 (2H, d,  $J = 8.0$  Hz, H-5, 9), 3.46 (1H, m, H-2), 2.98, 2.73 (2H, m, H-3); <sup>13</sup>C NMR (DMSO + D<sub>2</sub>O, 100 MHz): 171.96 (CO), 156.45 (C-7), 130.70 (C-6, 8), 116.32 (C-5, 9), 56.45 (C-2), 36.35 (C-3). Its IR, NMR data were consistent with those of L-tyrosine [14].

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